

Cobalt transport in a cobalt-resistant strain of *Neurospora crassa*

G. VENKATESWERLU and K. SIVARAMA SASTRY

Department of Biochemistry, Osmania University, Hyderabad 500 007

MS received 18 April 1979; revised 7 September 1979

Abstract. Uptake of Co^{2+} by cobalt-resistant strain is dependent on Co^{2+} concentration in the medium and is linear with time. The uptake is unaffected by metabolic inhibitors and decreased at low pH values. The uptake is independent of temperature in the range 0–40° C. The transport system is a passive diffusion process, unlike in the parent wild type strain where it is energy-dependent. It is possible that Mg^{2+} transport system is not involved in Co^{2+} transport in this strain, since the Co^{2+} uptake is not suppressed by Mg^{2+} as in the parent strain.

Keywords. *Neurospora crassa*; cobalt-transport; cobalt-resistant strain.

Introduction

A cobalt-resistant strain of *Neurospora crassa* isolated earlier was 10-fold more tolerant to cobalt and nickel; the mechanism of resistance was found not to involve the development of any permeability barrier to toxic metal ions (Venkateswerlu and Sivarama Sastry, 1973). Studies on the transport mechanism of Co^{2+} in the sensitive parent strain *N. crassa* Em 5297a revealed that a major part of the Co^{2+} uptake is energy-dependent and possibly mediated by the magnesium transport system (Venkateswerlu and Sivarama Sastry, 1970). Similar transport systems are known to exist in bacteria (Webb, 1970; Nelson and Kennedy, 1971, 1972). We report herein some features of Co^{2+} transport in a cobalt-resistant strain.

Materials and methods

Organisms, maintenance and growth

A cobalt-resistant strain of *N. crassa* was employed in these studies. The details of isolation of this strain, growth conditions and uptake have been described earlier (Venkateswerlu and Sivarama Sastry, 1970, 1973). All the uptake experiments were conducted using preformed mycelia of the cobalt-resistant strain. Co^{2+} concentrations are per 10 ml of medium.

Results

Kinetics of Co^{2+} uptake

The uptake of Co^{2+} by the cobalt-resistant strain as a function of time was first studied at three concentrations of Co^{2+} . It is evident that Co^{2+} uptake increases

linearly with Co^{2+} concentration in the medium (figure 1). The rate of uptake is $0.37 \mu\text{mol}$ ($22 \mu\text{g}$) $\text{Co}^{2+}/\text{h}/100 \text{ mg}$ dry weight of mycelia. Extrapolation of uptake to zero time indicates that surface binding in the cobalt-resistant strain is dependent on external cobalt concentration. As the cobalt concentration in the medium increases from 34 to $102 \mu\text{mol}$, the surface binding increases from 0.56 to about $1.6 \mu\text{mol}$ of cobalt/ 100 mg dry wt; this represents 23 and 47% , respectively of the total uptake since the rate of uptake of Co^{2+} is $0.37 \mu\text{mol}/100 \text{ mg}$ dry wt of mycelia per h, these values apparently represent a rapid initial binding of Co^{2+} by the cobalt resistant strain.

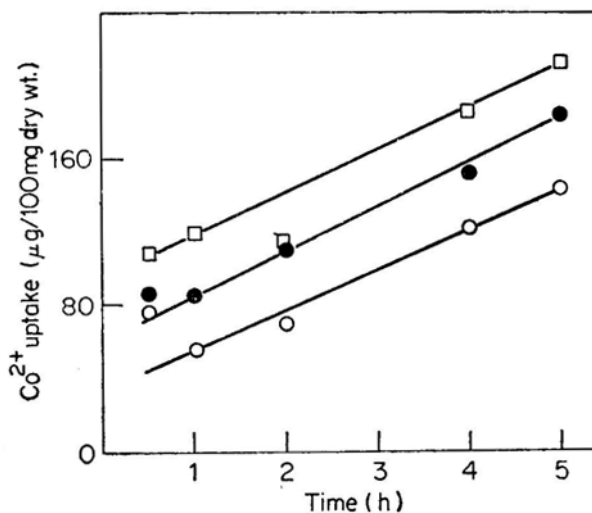


Figure 1. Rate of uptake of Co^{2+} by *N. crassa* cobalt-resistant strain. Preformed mycelia (72 h old) were incubated with varying concentrations of Co^{2+} for different time periods. (O) $34 \mu\text{mol}$ Co^{2+} ; (●) $68 \mu\text{mol}$ Co^{2+} ; (□) $102 \mu\text{mol}$ $\text{Co}^{2+}/10 \text{ ml}$ medium.

Also short term uptake of Co^{2+} by mycelia of the cobalt-resistant strain was studied as a function of Co^{2+} concentration for 10 and 20 min respectively. From the data of figure 2, it is evident that initial uptake is quite rapid, in contrast with the slower overall uptake, and is largely completed in 10 min. The zero-time uptake values obtained from figure 1, quite closely agree with these values, and support the conclusion that 23–47% of the Co^{2+} taken up at 34 – $102 \mu\text{mol}$ of Co^{2+} in the medium is surface bound.

Co^{2+} uptake by the cobalt-resistant strain has also been studied as a function of concentration from 0 – $204 \mu\text{mol}$ Co^{2+} in the incubation medium, using an incubation period of 3 h. It can be seen from figure 3, that the uptake increases rapidly upto a concentration of $3.39 \mu\text{mol}$ of Co^{2+} and thereafter, uptake increases much more slowly but nearly linearly with Co^{2+} concentration. It is interesting to note that, unlike in the parent strain, where uptake reaches nearly maximal value around $34 \mu\text{mol}$ of Co^{2+} in 2h (Venkateswerlu and Sivarama Sastry, 1970) the saturation of uptake does not occur in the cobalt-resistant strain upto the highest concentration of Co^{2+} tested in these experiments. However, when incubation with Co^{2+} is prolonged for 24 h (figure 3), there is a much higher uptake of Co^{2+}

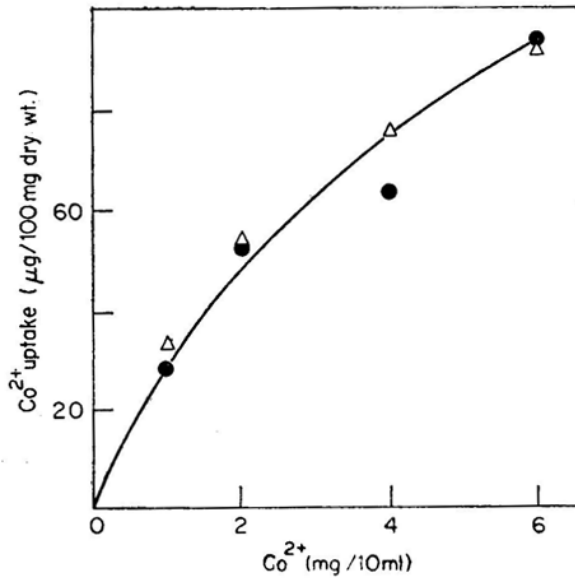


Figure 2. Short-term uptake of Co²⁺ by *N. crassa* cobalt-resistant strain. Preformed mycelia (72 h old) were allowed to take up Co²⁺ for 10 min (●) and 20 min (Δ) from medium containing various concentrations of Co²⁺.

and under these conditions of Co²⁺ concentration-dependent saturation is possible. This implies that in contrast with the parent strain, despite quantitatively larger uptake, saturation with Co²⁺ in the cobalt-resistant strain is attained much more slowly.

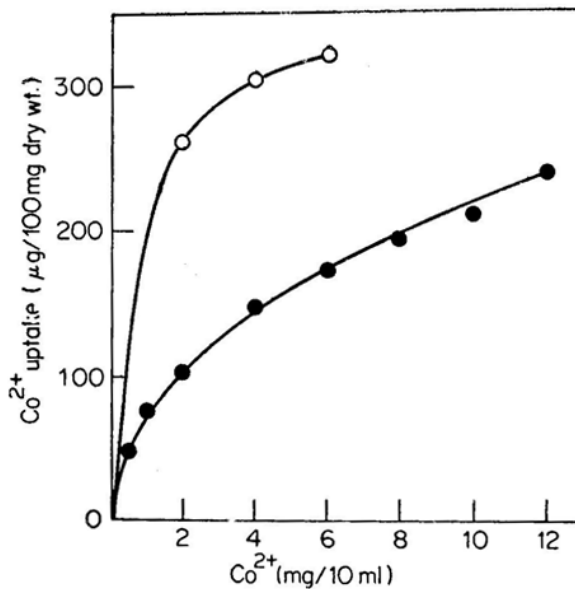


Figure 3. Dependence of Co²⁺ uptake by *N. crassa* cobalt-resistant strain on Co²⁺ concentration in the medium. Uptake by *N. crassa* cobalt-resistant strain on Co²⁺ concentration in the medium. Preformed mycelia (72h old) were incubated for 3h (●) or 24h (○) in media containing various concentrations of Co²⁺.

Effect of pH on Co²⁺ uptake

The effect of pH on Co²⁺ uptake by cobalt-resistant strain was also studied as described earlier (Venkateswerlu and Sivarama Sastry, 1970). The uptake was reduced below pH 4.0 and was unaffected in the range of pH 4–6. Decreased CO²⁺ uptake below pH 4.0 is evidently due to competition with H⁺ ions. It would appear therefore that there is no pH optimum for Co²⁺ uptake in the cobalt-resistant strain.

Effect of temperature on Co²⁺ uptake

Co²⁺ uptake by the cobalt-resistant strain has been studied in the temperature range of 050° C, and the temperature has little influence on Co²⁺ uptake in the range 040° C. However, a large increase in Co²⁺ uptake occurs at 50° C, 15 μmol of (885 μg) Co²⁺/100 mg dry weight in contrast to 2.63 μmol of Co²⁺/100 mg dry weight of mycelia in 3 h which possibly indicate nonspecific binding by thermally killed mycelial material.

Effect of metabolic inhibitors on Co²⁺ uptake

Intracellular uptake of Co²⁺ by the parent strain can be inhibited by 2,4-dinitrophenol, sodium azide and fluoride (Venkateswerlu and Sivarama Sastry, 1970). The results presented in table 1 indicate that none of the metabolic inhibitors tested significantly depress Co²⁺ uptake by the cobalt-resistant strain, maximal inhibition rarely exceeding 20% even at high concentration of these inhibitors.

Table 1. Effect of metabolic inhibitors on cobalt uptake by cobalt-resistant strain.

Inhibitor	Concentration (mM)	Co ²⁺ uptake (μmol/100 mg dry wt)	% inhibition
Control	..	3.60	0
Potassium fluoride	5	3.58	0
	10	3.01	17
Sodium azide	1	3.87	0
	2	2.95	18
2,4-Dinitrophenol	1	3.53	0
	2	3.05	15

Three day old mycelia were washed and incubated for 3 h with 102 μmol of cobalt/10 ml, in the presence of the inhibitor.

Effect of Mg²⁺ on Co²⁺ uptake

Excess Mg²⁺ in the medium is known to reverse the growth inhibition due to Co²⁺ toxicity in the parent strain by suppressing uptake of Co²⁺ by the mold (Sivarama

Sastry *et al.*, 1962); Mg²⁺ also suppresses Co²⁺ uptake by preformed mycelia of the parent strain (Venkateswerlu and Sivarama Sastry, 1970). In growth experiments, a similar phenomenon, was observed in the cobalt-resistant strain as well (Venkateswerlu and Sivarama Sastry, 1973). Hence, the effect of Mg²⁺ on Co²⁺ uptake by preformed mycelia of cobalt-resistant strain has been examined. The data presented in figure 4 indicate that Mg²⁺, when present in a two-fold excess (by wt), is unable to inhibit the Co²⁺ uptake, while this concentration of Mg²⁺ results in the complete reversal of growth inhibition caused by Co²⁺ (Venkateswerlu and Sivarama Sastry, 1973). It would seem that, in this respect the cobalt-resistant strain differs from the parental strain. Since in growth experiments, Mg²⁺ does however suppress overall uptake of Co²⁺, it is evident that Mg²⁺ does inhibit Co²⁺ uptake in the cobalt-resistant strain but less significantly.

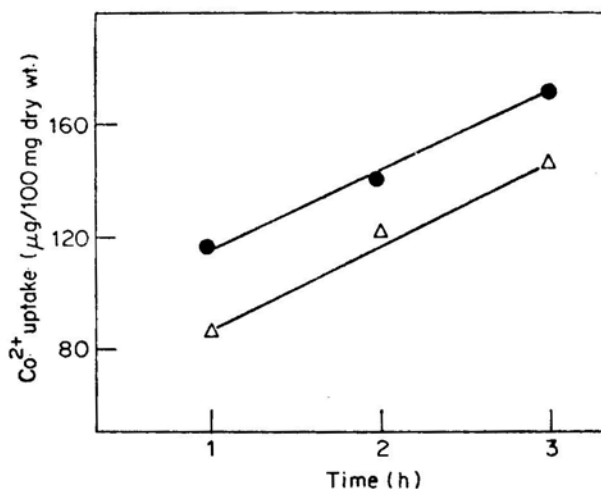


Figure 4. Effect of Mg²⁺ on Co²⁺ uptake by *N. crassa* cobalt-resistant strain. Preformed mycelia (72 h old) were incubated with 102 µmol of Co²⁺ for different time periods in presence of 0.82 µmol of Mg²⁺ (●) and 494 µmol of Mg²⁺ (Δ) in 10 ml of medium.

Effect of EDTA on Co²⁺ uptake by preformed mycelia

It is evident that approximately 25–50% of the total Co²⁺ taken up by preformed mycelia of cobalt-resistant strain is bound at the surface. When preformed mycelia of the parent strain of *N. crassa* saturated with Co²⁺ were floated in a solution of 1 mM EDTA, rapid removal of (within 30 min) 20% of the Co²⁺ taken up was observed. A much slower release of Co²⁺ (another 10%) was observed within the next 2.5 h after which EDTA has little effect (Venkateswerlu and Sivarama Sastry, 1970). Similar experiments conducted in the present study with the cobalt-resistant strain revealed that around 80% of the total Co²⁺ taken up is leached out by EDTA in 7 h (see figure 5), and that the process levels off at this stage.

The data in figure 5 also indicate that leaching of Co²⁺ by EDTA is slow and progressive. It is probable that the slow rate of this efflux is due to a slow influx rate of EDTA and/or a slow efflux rate for a Co²⁺ EDTA complex.

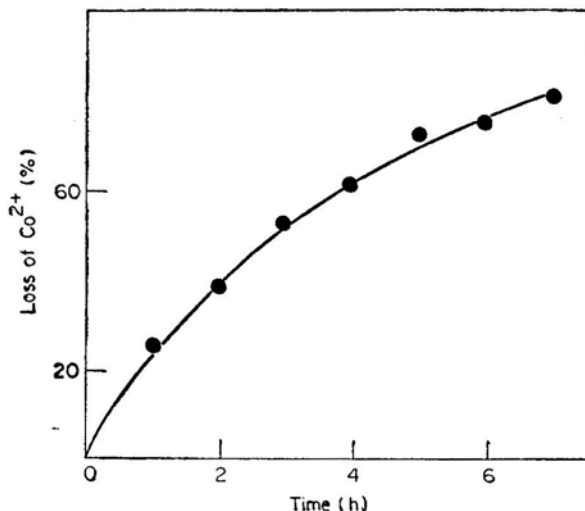


Figure 5. Effect of EDTA on Co^{2+} taken up by *N. crassa* cobalt-resistant strain. Preformed mycelia (72 h old) were incubated with $102 \mu\text{mol Co}^{2+}/10\text{ml}$ for 3 h, washed and resuspended in medium containing 1mM EDTA for different times.

In this context it is significant to note that the part of uptake which is sensitive to inhibition by metabolic inhibitors (18.5% of total, see table 1) is resistant to removal by EDTA.

Discussion

In contrast to our earlier observations of an energy-dependent Co^{2+} uptake which is mediated through the Mg^{2+} transport system in the parent strain of *N. crassa* (Venkateswerlu and Sivarama Sastry, 1970), Co^{2+} transport system exhibits different properties in the cobalt-resistant strain. This system manifests a rapid initial uptake of Co^{2+} (23% at $34 \mu\text{mol}$ of Co^{2+} and 47% at $102 \mu\text{mol}$ of Co^{2+}). This suggests that an equilibrium exists between free and cell surface bound Co^{2+} in the cobalt-resistant strain. Unlike in the parent strain, the time taken to saturate the uptake system is very long in the cobalt-resistant strain. The concentration of Co^{2+} required for this is also very high as compared to the parent strain (Venkateswerlu and Sivarama Sastry, 1970).

The uptake of Co^{2+} in cobalt-resistant strain is not temperature-dependent. Similar instances are known in bacteria (Webb, 1970; Silver and Kralovic, 1969). Webb (1970) showed that the energy-independent uptake of Co^{2+} in metal-resistant strains of bacteria is invariant between $0\text{--}28^\circ\text{C}$. Ca^{2+} uptake by *E. coli* is unchanged with temperature between $25\text{--}45^\circ\text{C}$ (Silver and Kralovic, 1969). Also, metabolic inhibitors do not inhibit Co^{2+} uptake significantly in the cobalt-resistant strain. These evidences favour the existence of an energy-independent Co^{2+} transport system in the cobalt-resistant strain. However, whether this altered transport system for Co^{2+} has any relation with the acquisition of resistance to Co^{2+} is not clear.

Another difference indicated by these results pertains to the nature of intracellular Co²⁺. In the parent strain, 30% of bound Co²⁺ is removed by EDTA from the preformed mycelia that accumulated Co²⁺. However, this is not the case with the cobalt-resistant strain incubated with Co²⁺. These observations indicate that Co²⁺ is relatively more loosely bound intracellularly in the Cobalt-resistant strain or reflects the differences in the uptake of EDTA.

Apparently, the greater extracellular concentration of Co²⁺ (102 μ mol) required for 50% growth inhibition in the cobalt-resistant strain coupled with the nature of the slow diffusion process involved, results in an eventual build-up of Co²⁺ concentration in mycelia of cobalt-resistant strain. This is also evident from the difference in uptake observed between 3 h and 24 h uptake with preformed mycelia.

Acquisition of metal resistance is obviously not due to the development of a permeability barrier for Co²⁺ ions; however, metal uptake patterns at the cell-surface are very different in the cobalt-resistant strain as shown by effects of inhibitors, EDTA and the rather unusual aspects of Co²⁺-Mg²⁺ antagonism at the level of Co²⁺ uptake.

There are two probable mechanisms which could explain the present data. The first of these is that the metal transport system in *N. crassa* which transports Mg²⁺ and Co²⁺ has widely different specificities in the cobalt-resistant strain. On this basis, competitive effects would be quantitatively different depending upon the nature of the change in affinity. Alternatively, the transport system is no longer able to mediate Co²⁺ uptake, with the consequence that uptake is restricted to passive diffusion. This implies that the Mg²⁺-Co²⁺ antagonism is much less pronounced at the cell surface in the cobalt-resistant strain and is restricted to competition in passive diffusion. If Mg²⁺ transport is predominantly mediated by a specific transport system and this system also usually transports Co²⁺, it is easy to see why Mg²⁺ cannot powerfully suppress Co²⁺ uptake in the cobalt-resistant strain.

Acknowledgement

G. Venkateswerlu thanks the Department of Atomic Energy, Government of India, for the award of a Research Fellowship.

References

- Nelson, D. L. and Kennedy, E. P. (1971) *J. Biol. Chem.*, **246**, 3042.
- Nelson, D. L. and Kennedy, E. P. (1972) *Proc. Natl. Acad. Sci. USA*, **69**, 1091.
- Silver, S. and Kralovic, M. L. (1969) *Biochem. Biophys. Res. Commun.*, **34**, 640.
- Sivarama Sastry, K., Adiga, P. R., Venkatasubramanyam, V. and Sarma, P. S. (1962) *Biochem. J.*, **85**, 486.
- Venkateswerlu, G. and Sivarama Sastry, K. (1970) *Biochem. J.*, **118**, 197.
- Venkateswerlu, G. and Sivarama Sastry, K. (1973) *Biochem. J.*, **132**, 673.
- Webb, M. (1970) *Biochim. Biophys. Acta*, **222**, 440.