# Influence of biochemical parameters of host plants on the biology of *Dialeurodes vulgaris* Singh (Aleyrodidae: Homoptera)

### **R** SUNDARARAJ and **B** V DAVID

Fredrick Institute of Plant Protection and Toxicology, Padappai 601 301, India MS received 16 February 1990

Abstract. The influence of biochemical parameters of the host plants viz. Jasminum multiflorum Andr., Jasminum sambac Ait., Jasminum grandiflorum Linn., Jasminum auriculatum Vahl., Jasminum pubescens Willd. and Jasminum flexile Vahl. on the biology of Dialeurodes vulgaris was studied. The results indicate remarkable variation in the biochemical parameters of host plants which affected significantly the biology of the insect. The per cent survival and growth index have been considerably low in Jasminum pubescens, Jasminum auriculatum and Jasminum grandiflorum and the same trend has been noticed with regard to fecundity and longevity also. The most preferred hosts have been Jasminum sambac and Jasminum multiflorum. It is observed that high amount of phenol and excessive accumulation of sugars in host plants make them nonpreferred while high amount of amino acids makes the plants more preferred by Dialeurodes vulgaris.

Keywords. Dialeurodes vulgaris; jasmine; host preference.

#### 1. Introduction

Several insects infest jasmine (Jasminum sambac) crop in south India (David 1958). Among them the jasmine whitefly Dialeurodes vulgaris Singh has been noticed to be endemic on the cultivars of jasmine in Tamil Nadu. There are 7 cultivars of jasmine belonging to 6 species of the genus Jasminum and all of them are infested by D. vulgaris. The present study reports on the quantitative levels of certain biochemical parameters of these cultivars and their influence on the biology of the whitefly on them.

#### 2. Materials and methods

Seven cultivars of jasmine viz. Kundumalli (J. multiflorum Andr.), Maduramalli (J. sambac Ait.), Adukkumalli (J. sambac Ait.), Jathimalli (J. grandiflorum Linn.), Virijathi (J. auriculatum Vahl.), Kakkadamalli (J. pubescens Willd.) and Nithyamalli (J. flexile Vahl.) were raised in polythene bags with soils. Transparent plastic cages  $(8 \times 6 \text{ cm})$  with screw cap at one end and the other end covered with muslin cloth were prepared. A part of the host plant was enclosed inside the cage. Whiteflies from the stock culture were collected using an aspirator and released into the cages for oviposition on the leaves. After 24 h the whiteflies and the cages were removed and observations recorded for incubation period, nymphal period, total developmental period, percentage survival, growth index, longevity of male and female and fecundity. There were 5 replications for each study.

Leaf samples of the host plants were simultaneously analysed for quantitative estimation of biochemical parameters. Leaf extracts were obtained using 80% ethanol. The reducing sugars were determined first. Subsequently the nonreducing sugars in the extract were hydrolysed to reducing sugars and the total sugars were

## 138 R Sundararaj and B V David

estimated by Nelson's (1944) method. By subtracting the reducing sugars from the total sugars, the nonreducing sugars were estimated and expressed as glucose equivalents. Free amino acids, protein and phenol were estimated following the methods, respectively of Moore and Steins (1948), Lowry *et al* (1951) and Bray and Thorpe (1954) (Outlined by Mahadevan and Sridhar 1982).

## 3. Results and discussion

The data presented in table 1 clearly indicate that the total developmental period have been considerably short whereas the per cent survival and growth index have been appreciably high in *J. sambac* and *J. multiflorum* followed by *J. flexile*, and the same trend has been noticed with regard to fecundity also. Hence the most preferred hosts have been *J. sambac* and *J. multiflorum* whereas *J. pubescens* proved to be the least preferred one.

# 3.1 Influence of biochemical parameters of host plants on D. vulgaris

The data on reducing sugar, non reducing sugar, total sugar, free amino acids, protein and phenol for the different species of *Jasminum* are given in table 2. The most unfavourable host *J. pubescens* (Kakkadamalli) contains the highest amount of phenol (21-238 mg/g) followed by *J. grandiflorum* (Jathimalli) which contains 9,000 mg/g. When phenol is sufficiently synthesised it combines with the sugars obtained as primary product of photosynthesis to form tannins. The tannins combine irreversibly with the available protein and form the indigestible complexes which result in the nutritional deficiency in the host plants which in turn affects the percentage of infestation and host preference (Raman and Ananthakrishnan 1986). On the contrary *J. auriculatum* (Virijathi) recorded lowest amount of phenol

|                                 | Incuba-<br>tion | Nymphal<br>period | Total<br>develop-<br>mental period | Survival          | Growth | Longevity<br>(days) |        |           |
|---------------------------------|-----------------|-------------------|------------------------------------|-------------------|--------|---------------------|--------|-----------|
| Host                            | (days)          | (days)            | (days)                             | (%)               | index  | Male                | Female | Fecundity |
| Kundumalli<br>(J. multiflorum)  | 8.06            | 24.68             | 32.74                              | 71-26             | 2.18   | 11.10               | 12.93  | 82.00     |
| Maduramalli<br>(J. sambac)      | 8-13            | 24.45             | 32.58                              | <del>68</del> ·78 | 2.11   | 10-57               | 12.10  | 75-33     |
| Adukkumalli<br>(J. sambac)      | 8·26            | 24.53             | 32.79                              | 72-17             | 2.20   | 9.77                | 11-47  | 71-67     |
| Jathimalli<br>(J. grandiflorum) | 8.32            | 26-32             | 34.63                              | 54-26             | 1-57   | 6.63                | 7·40   | 46.63     |
| Virijathi<br>(J. auriculatum)   | 8.62            | 27.00             | 35-63                              | 47-91             | 1.36   | 6-26                | 6.57   | 36.00     |
| Kakkadamalli<br>(J. pubescens)  | 8-38            | 26-60             | 34.97                              | 39-26             | 1.12   | 4.57                | 5.63   | 26.33     |
| Nithyamalli<br>(J. flexile)     | 8.84            | 24.40             | 33·24                              | 63-83             | 1-91   | 9.83                | 10-13  | 68·67     |
| $\overline{\text{CD}(P=0.05)}$  | NS              | 1-46              | 1.15                               | 5.71              | 0.20   | 1.48                | 1.45   | 1007      |

Table 1. Influence of hosts on the developmental biology of D. vulgaris.

| <u></u>                                |                   | Non               |                | Free           |         |        |
|--|-------------------|-------------------|----------------|----------------|---------|--------|
| Host                                   | Reducing<br>sugar | reducing<br>sugar | Total<br>sugar | amino<br>acids | Protein | Phenol |
| Kundumalli<br>(J. multiflorum)         | 32.565            | 23.495            | 55.727         | 2.512          | 22.248  | 4.694  |
| Maduramalli<br>(J. sambac)             | 33.689            | 25.869            | 58.557         | 2.776          | 19.748  | 6.492  |
| Adukkumalli<br>(J. sambac)             | 27.963            | 20-341            | 48.304         | 2.530          | 26.163  | 4-805  |
| Jathimalli<br>(J. grandiflorum)        | 22.141            | 10-315            | 32.456         | 2.319          | 22.443  | 9.000  |
| Virijathi<br>( <i>J. auriculatum</i> ) | 18.324            | 5.547             | 23.871         | 1.886          | 13.336  | 1.143  |
| Kakkadamalli<br>(J. pubescens)         | 48-383            | 7.721             | 56.103         | 2.093          | 23.808  | 21.238 |
| Nithyamalli<br>(J. flexile)            | 68·643            | 9.641             | 73-285         | 2.606          | 26.489  | 3.139  |
| $\overline{\text{CD}(P=0.05)}$         | 5.616             | 11.110            | 10.724         | 0.148          | 3.063   | 0.678  |

Table 2. Biochemical parameters\* of host plants of D. vulgaris.

\*Values in mg/g (mean of 3 replications).

(1.143 mg/g) and its non preference may be due to the low nutritive contents viz. reducing sugar (18.324 mg/g), non reducing sugar (5.547 mg/g), total soluble sugar (23.871 mg/g), free amino acids (1.886 mg/g) and protein (13.336 mg/g). House (1969) stated that in general all the phytophagous insects have quite similar qualitative nutritional requirements but it is the quantitative factor that plays a more decisive role in insect host plant relationship. The total soluble sugar was found to be the highest (73.285 mg/g) in J. flexile (Nithyamalli) which is moderately preferred. The antibiosis or non preference of the host plants was correlated with the excessive accumulation of total sugars (Ananthakrishnan 1986). The favourable hosts viz. J. sambac (Adukkumalli/Maduramalli) and J. multiflorum (Kundumalli) contain moderate amount of sugars and protein and low amount of phenol when compared with J. pubescens (Kakkadamalli) and J. grandiflorum (Jathimalli). The amount of free amino acids was higher in preferred hosts being 2.512 mg/g in J. multiflorum (Kundumalli), 2.776 mg/g in J. sambac (Maduramalli) and 2.530 mg/g in J. sambac (Adukkumalli) which is in agreement with the findings of David and Paul (1973). However, J. flexile (Nithyamalli) recorded higher amount of amino acid and its non preference may be due to excessive accumulation of sugars (73.285 mg/g).

#### Acknowledgements

The authors are thankful to Dr T N Ananthakrishnan, Entomology Research Institute, Loyola College, Madras for critically going through the manuscript. Thanks are due to Mr C Tamilselvan and Mr James Fredrick, for facilities.

#### References

Ananthakrishnan T N 1986 Nature of insect-plant interactions; in Dynamics of insect-plant interactions (eds) T N Ananthakrishnan and S Viswanathan (Madras: P and P Pvt Ltd.) pp 1-15 Bray H G and Thorpe W V 1954 Analysis of phenolic compounds of interest in metabolism; Meth. Biochem. Anal. 1 27-52

David S K 1958 Insects and mites affecting jasmine in the Madras state; Madras Agric. J. 45 146-150

David B V and Paul A V N 1973 Studies on resistance of castor to the whitefly Trialeurodes rara Singh I. Free amino acids; Madras Agric. J. 60 1499-1503

 House H L 1969 Effects of different proportions of nutrients on insects; Entomol. Exp. Appl. 12 651-669
Lowry O H, Rosebrough N J, Fan A L and Randall R J 1951 Protein measurement with the Folinphenol reagent; J. Biol. Chem. 193 265-275

- Moore S and Stein W H 1948 Photometric method for use in the chromatography of amino acids; J. Biol. Chem. 176 367-388
- Nelson N 1944 A photometric adaptation of the Somogyi method for the determination of glucose; J. Biol. Chem. 153 375-380
- Raman K and Ananthakrishnan T N 1986 Mechanisms of host plant selection in phytophagous insects; in *Dynamics of insect-plant interactions* (eds) T N Ananthakrishnan and S Viswanathan (Madras: P and P Pvt. Ltd.) pp 16-36